

Plant Biotechnology

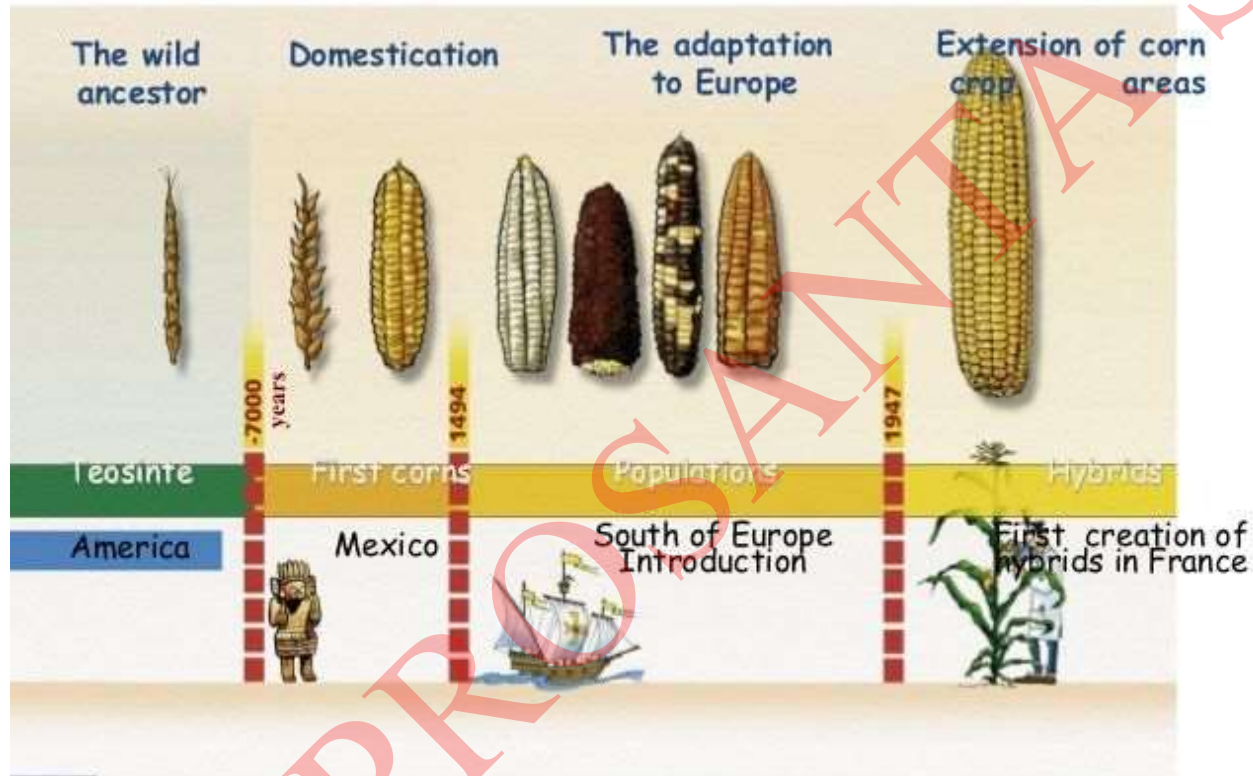
-Overview

Prosanta Saha

Domestication for crop improvement

- A process over thousands of years... Since human civilization began
- Mainly done through conventional breeding approaches

The evolution of maize (corn)



plant breeding and biotechnology contd...

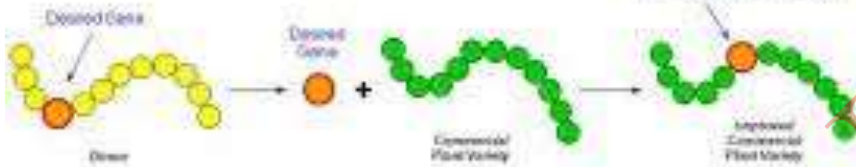
TRADITIONAL PLANT BREEDING

DNA is a strand of genes, much like a strand of pearls. Traditional plant breeding combines many genes at once.



PLANT BIOTECHNOLOGY

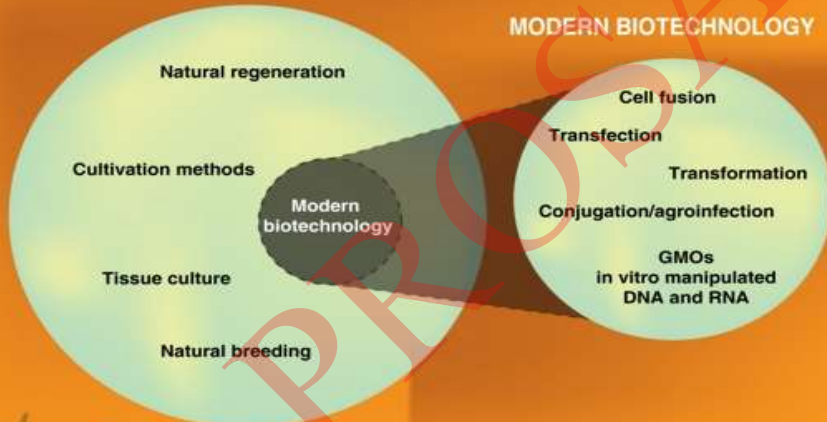
Using plant biotechnology, you can add a single gene to the strand.



Comparing Genetic Modification Techniques

	Conventional Breeding	Mutation breeding	Plant Biotechnology
Level	Whole organism	Molecule	Molecule
Precision	Thousands of genes	Unknown	Single gene
Certainty	Genetic change poorly characterized	Genetic change poorly characterized	Gene function well understood
Limits	Between species and genera	Not applicable	No limitations

Biotechnology



Genetic Engineering

The process of manipulating and transferring instructions carried by genes from one cell to another

Why do scientists want to change gene instructions?

- to produce needed chemicals**
- to carry out useful processes**
- to give an organism desired characteristics**

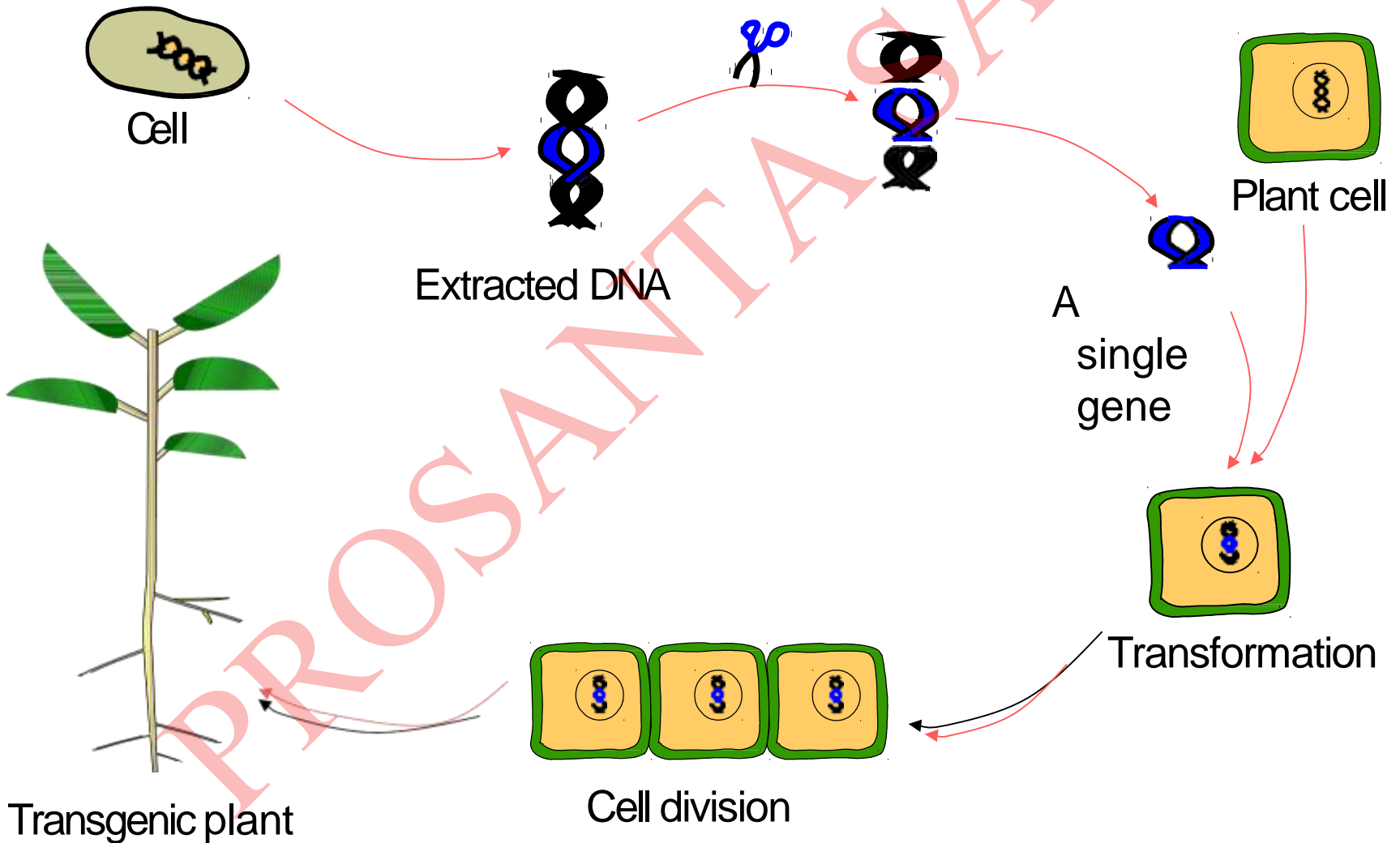
What About the Term Genetic Engineering?

Genetic engineering is the basic tool set of biotechnology

Genetic engineering involves:

- Isolating genes
- Modifying genes so they function better
- Preparing genes to be inserted into a new species
- Developing transgenics

Plant Genetic Engineering Process



Introducing the Gene - Requirements

STEPS

Create transformation cassette

Introduction of the gene

Selection of transformants

- **Prepare tissue for transformation**
 - Tissue must be capable of developing into normal plants
 - Leaf, germinating seed, immature embryos
- **Introduce DNA**
 - *Agrobacterium* or gene gun
 - **DNA delivery systems must be**
 - *Simple*
 - *Efficient and preferably inexpensive*
 - *The method must be available for use either because it is in the public domain or because it can be licensed*
- **Culture plant tissue**
 - Develop shoots & Roots
- **Screening of putative transformants**
 - Field test the plants
 - **System of choice depends on**
 - *the target plant*
 - *its regeneration system*

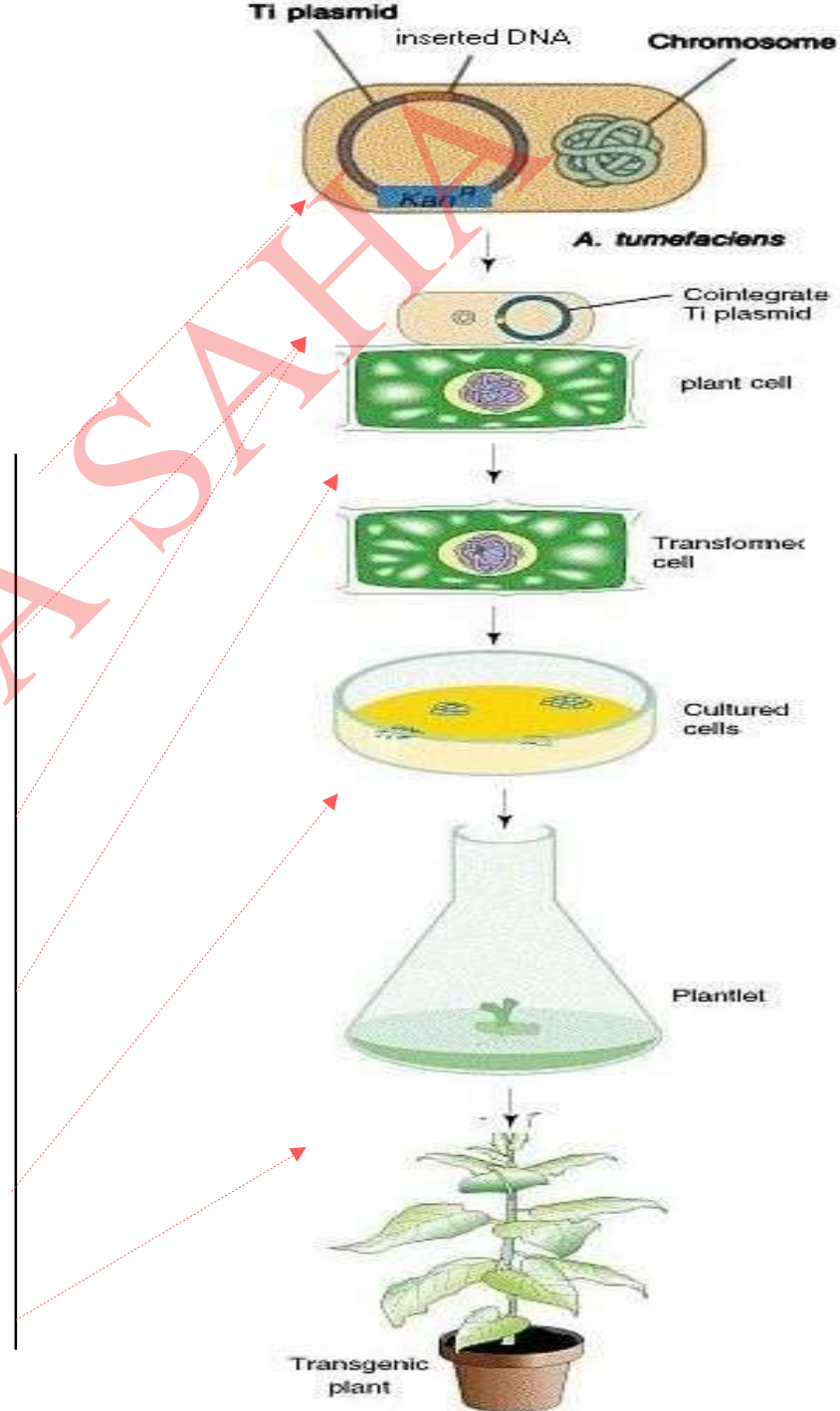
Isolate and clone gene of interest

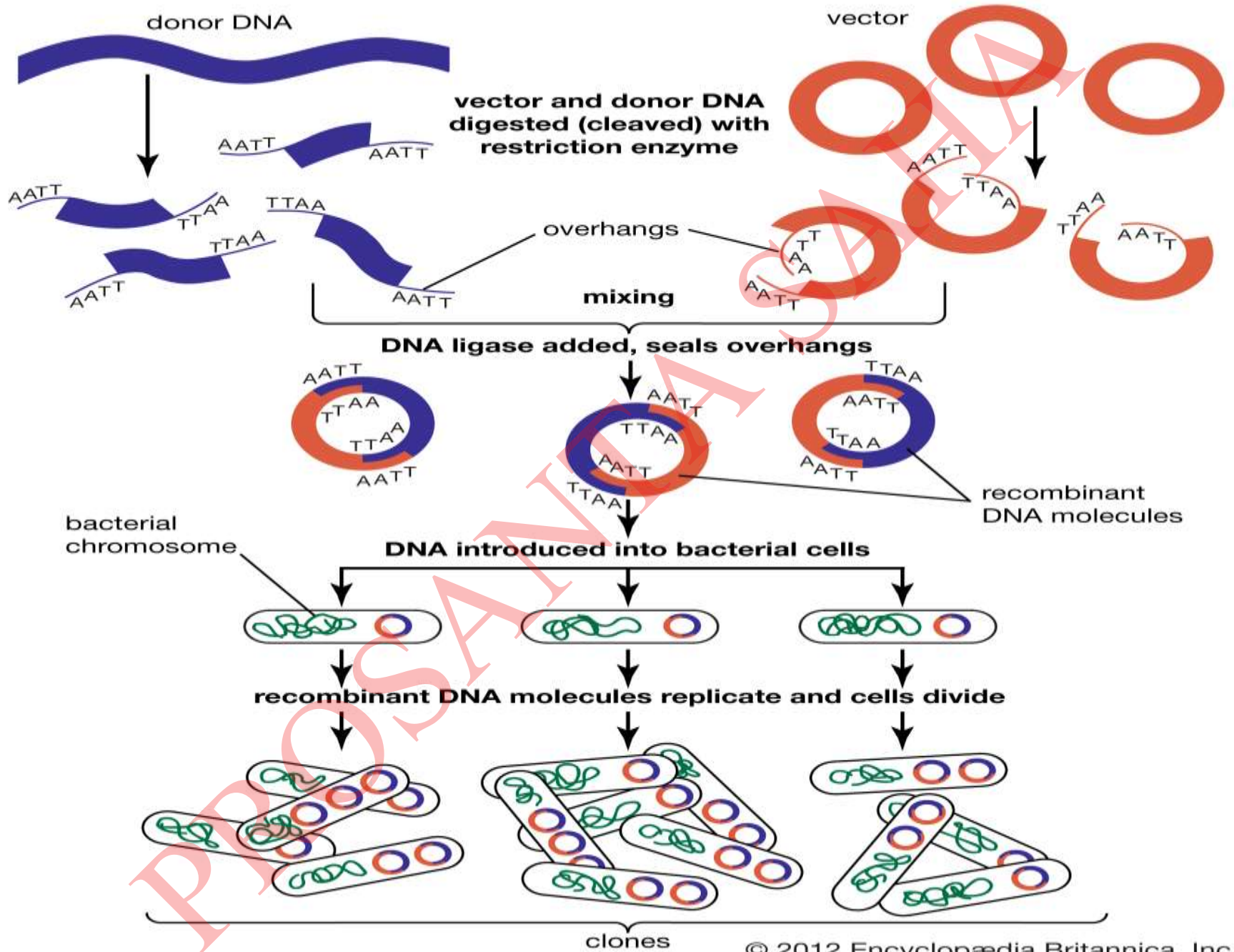
Add DNA segments to initiate or enhance gene expression

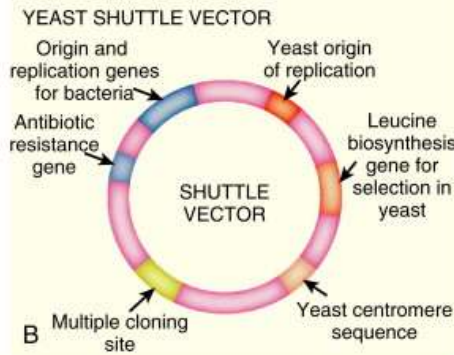
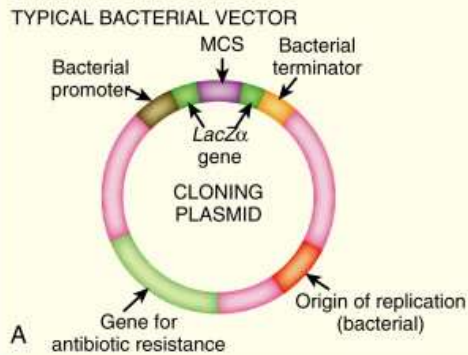
Add selectable markers Introduce gene construct into plant cells (transformation)

Select transformed cells or tissues

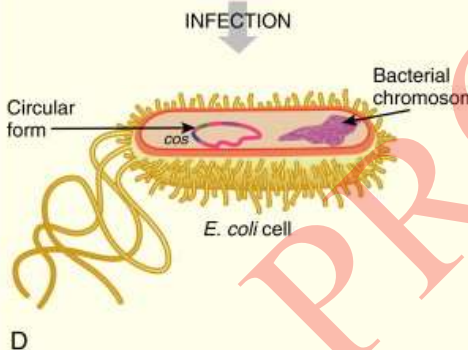
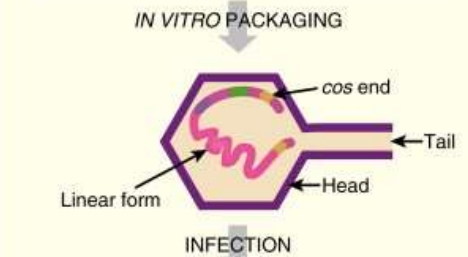
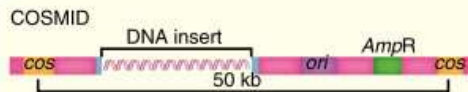
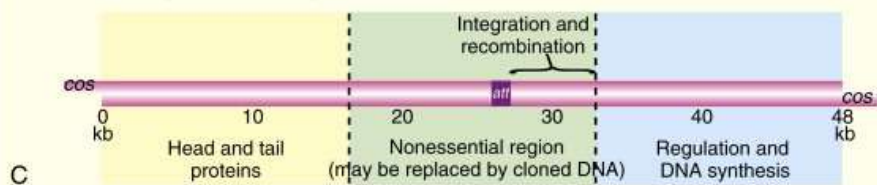
Regenerate whole plants



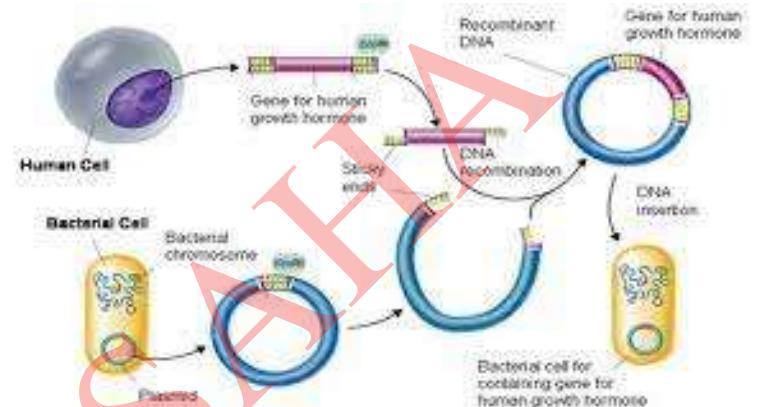
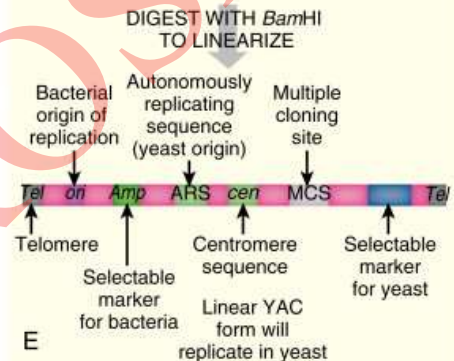
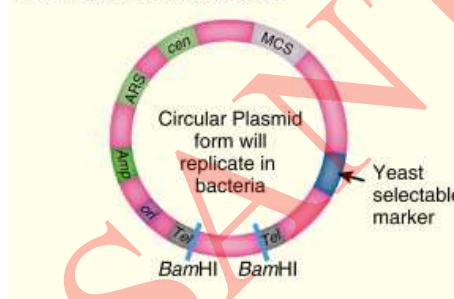




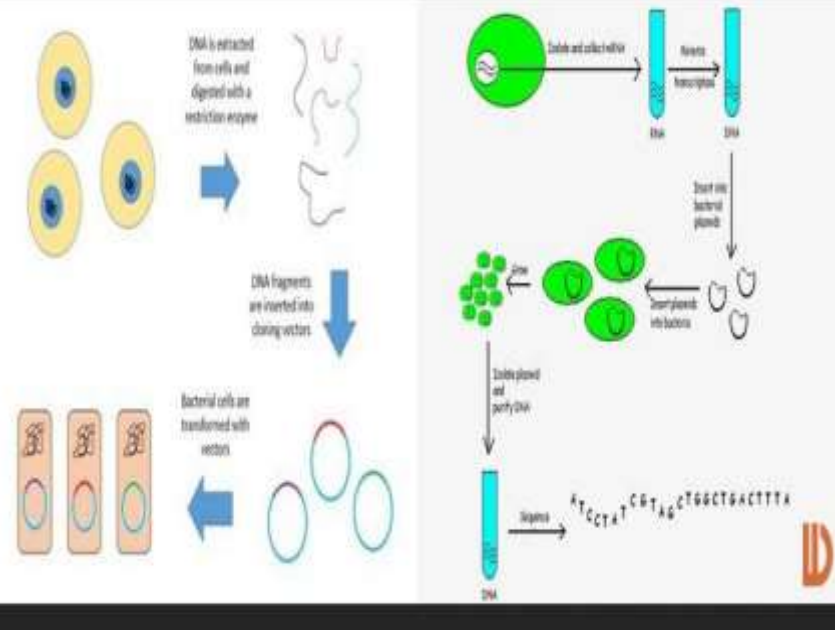
LAMBDA REPLACEMENT VECTOR



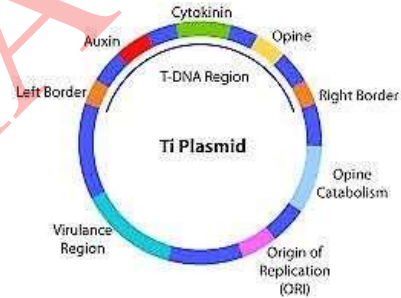
ARTIFICIAL CHROMOSOME



GENOMIC LIBRARY VS CDNA LIBRARY



Commonly used promoters



• **Constitutive promoter**

- **CaMV 35S**: suitable for expression of foreign genes in dicots:
- The maize **ubiquitin** promoter, also a constitutive promoter which
- drives strong expression of transgenes in monocots.

• **Organ/ tissue specific promoters**

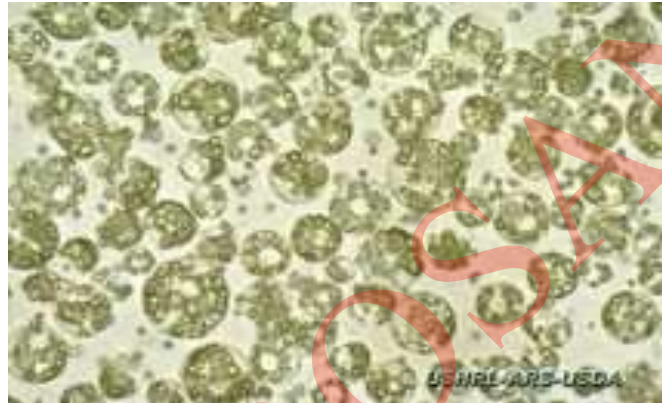
- **Vicilin** and **phytohemagglutinin, glutenin promoters** seed specific expression
- **a-amylase** promoter for expression in the aleurone of cereal grains;
- **Patatin** promoter for tuber specific expression in potatoes and the **RuBisCo** promoter for green tissuespecificity

Regeneration

We use tissue culture techniques to regenerate whole plants from single cells

getting a plant back from a single cell is important so that every cell has the new DNA

Plant tissue culture uses growth regulators and nutrients to regenerate plants *in vitro*



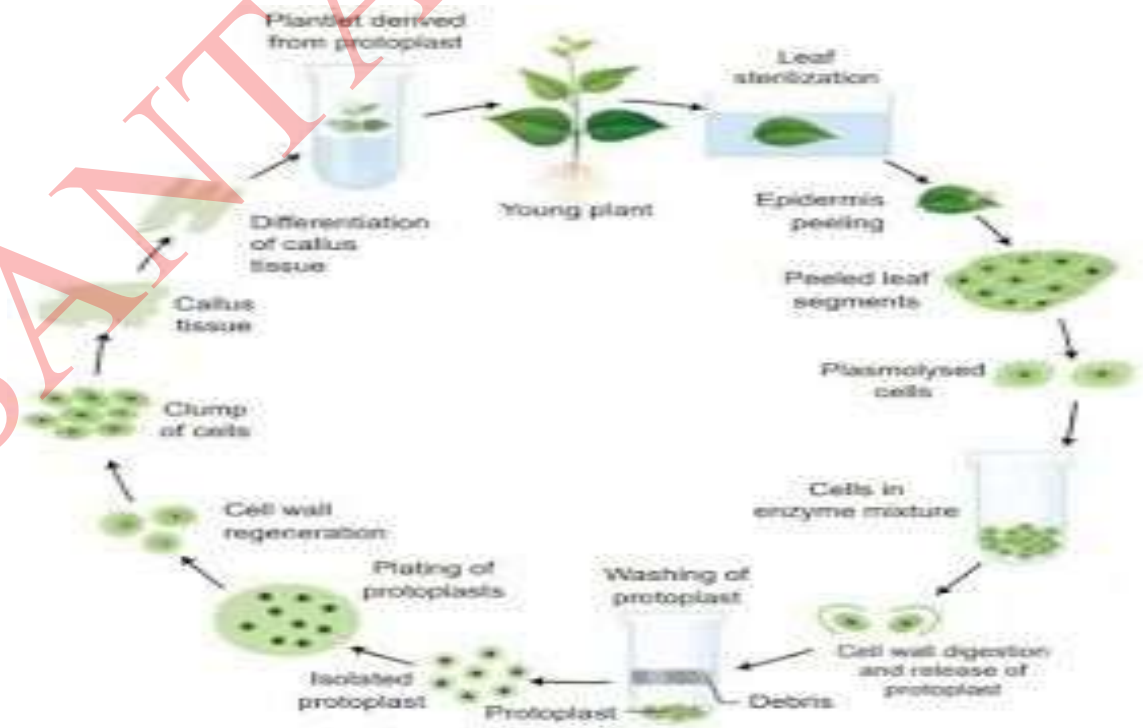
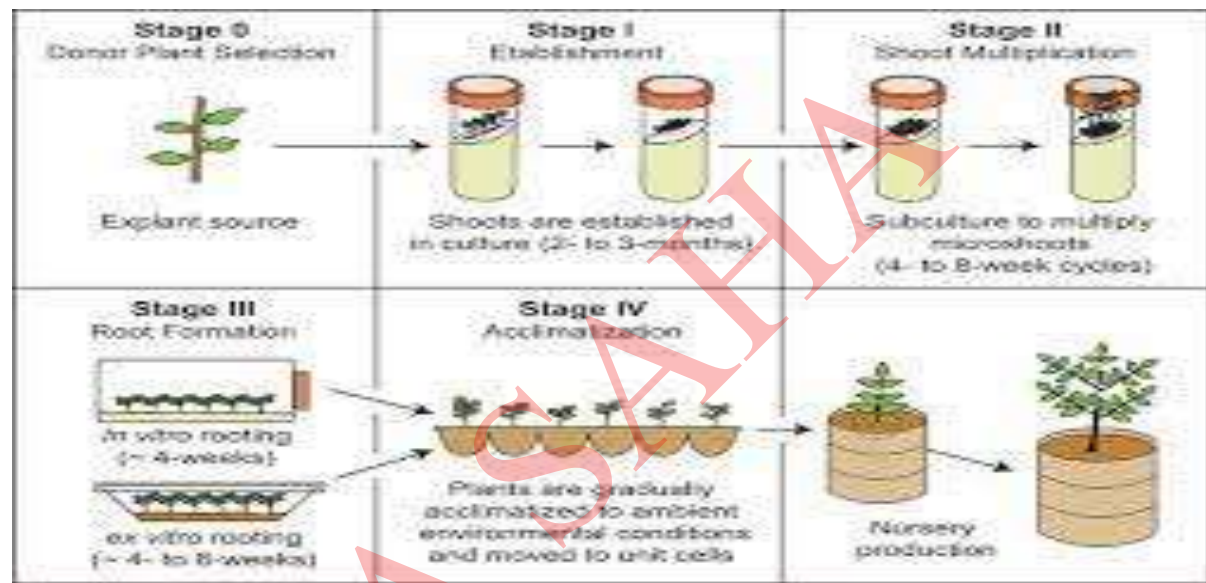
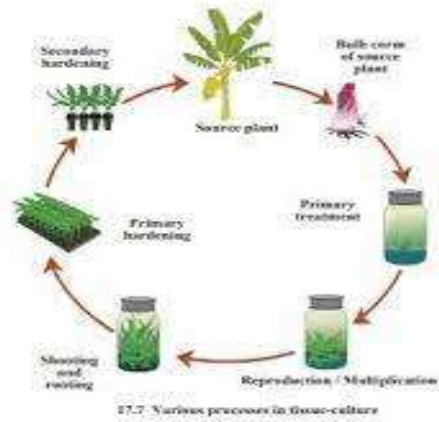


Figure 5.8: Protoplast Culture

Transformation methods

Indirect

Agrobacterium tumefaciens mediated, Virus mediated

Direct

Particle bombardment

Polyethylene glycol (PEG)-mediated protoplast transformation.

Electroporation Microinjection

Silicon Carbide Whiskers (SCW)

Method depends on plant type, cost, application

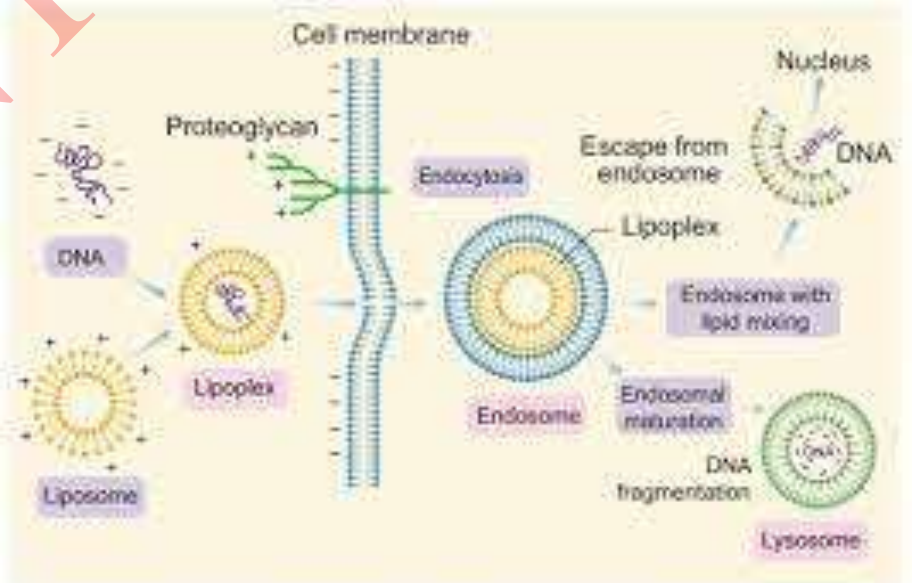
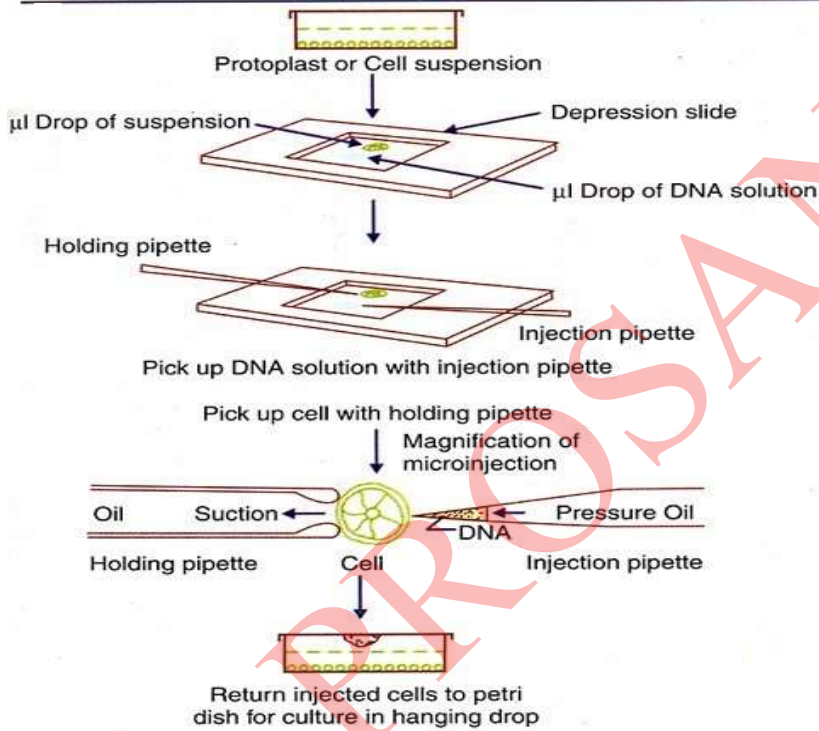
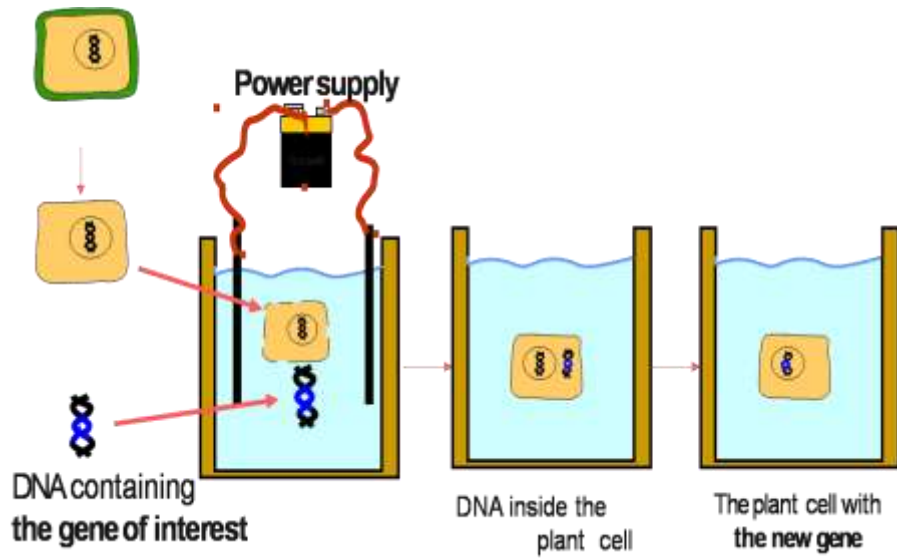
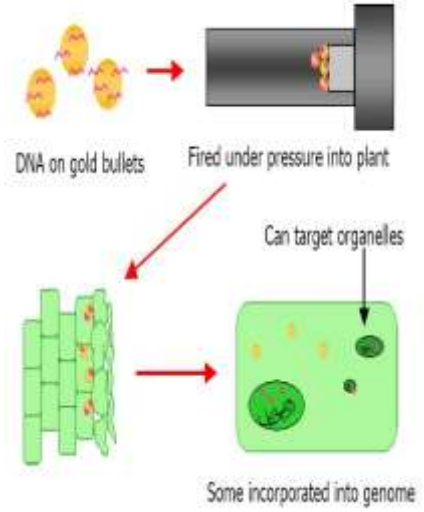
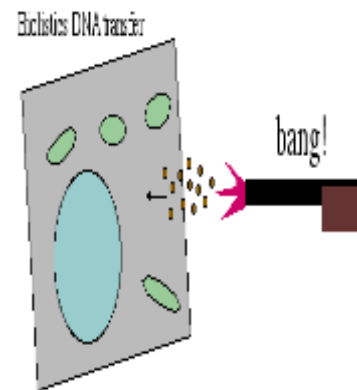
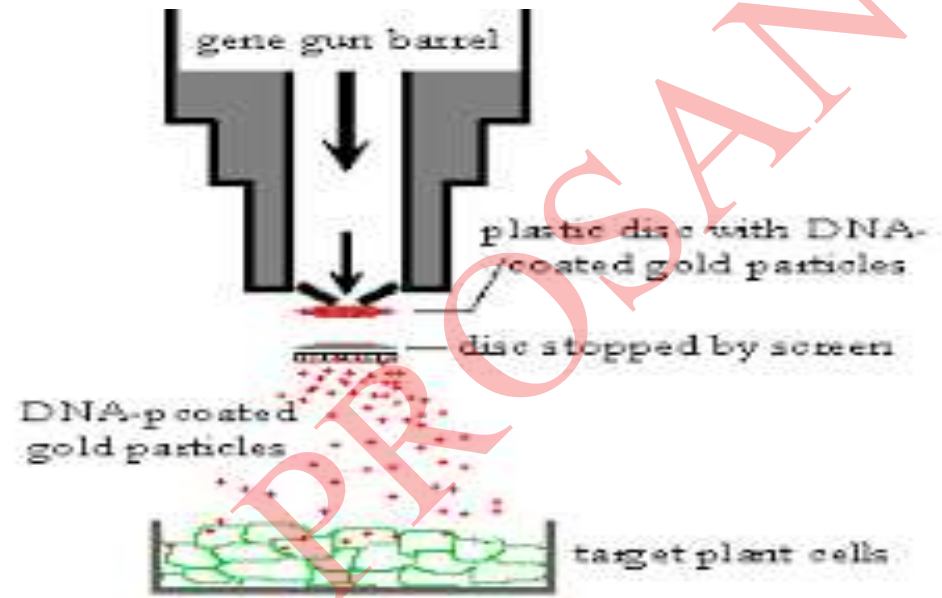
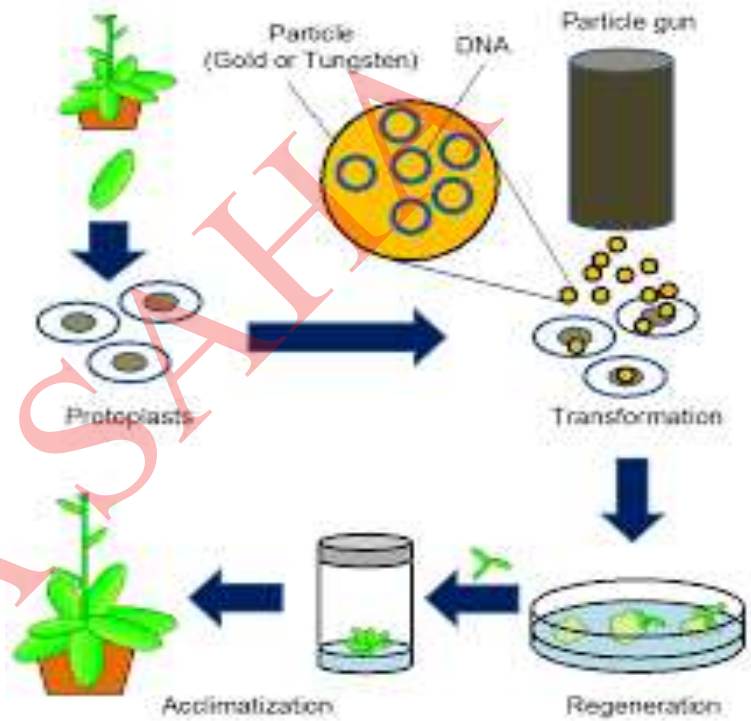
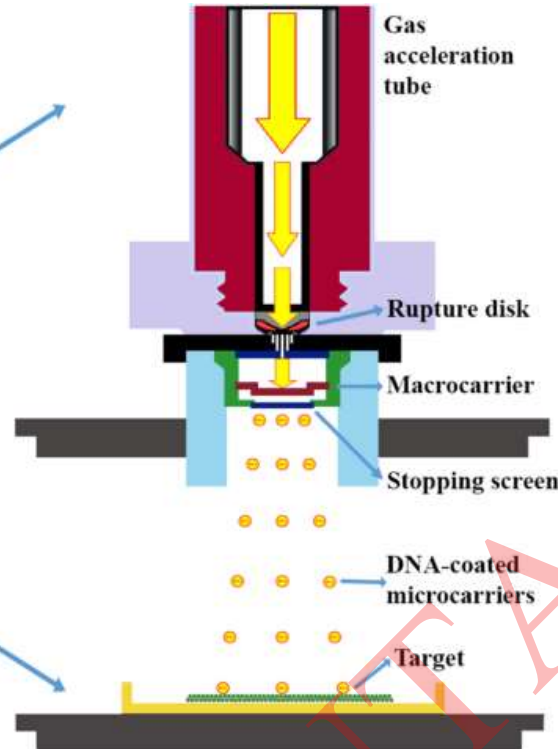
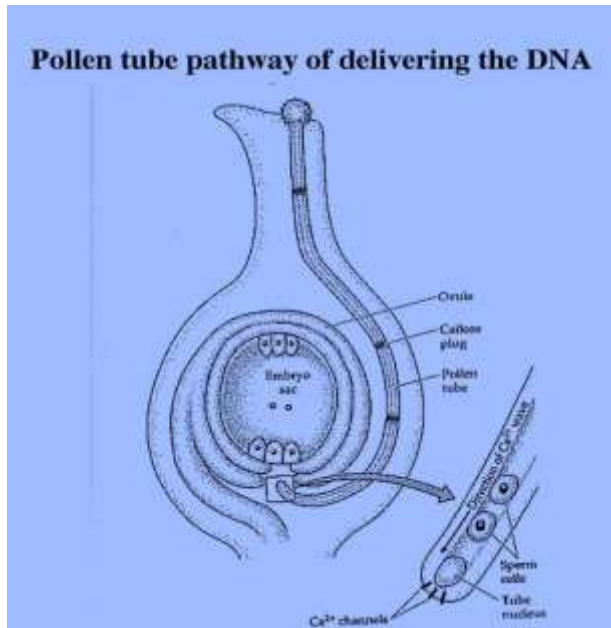


Figure 4.15: Liposome mediated method of Gene Transfer



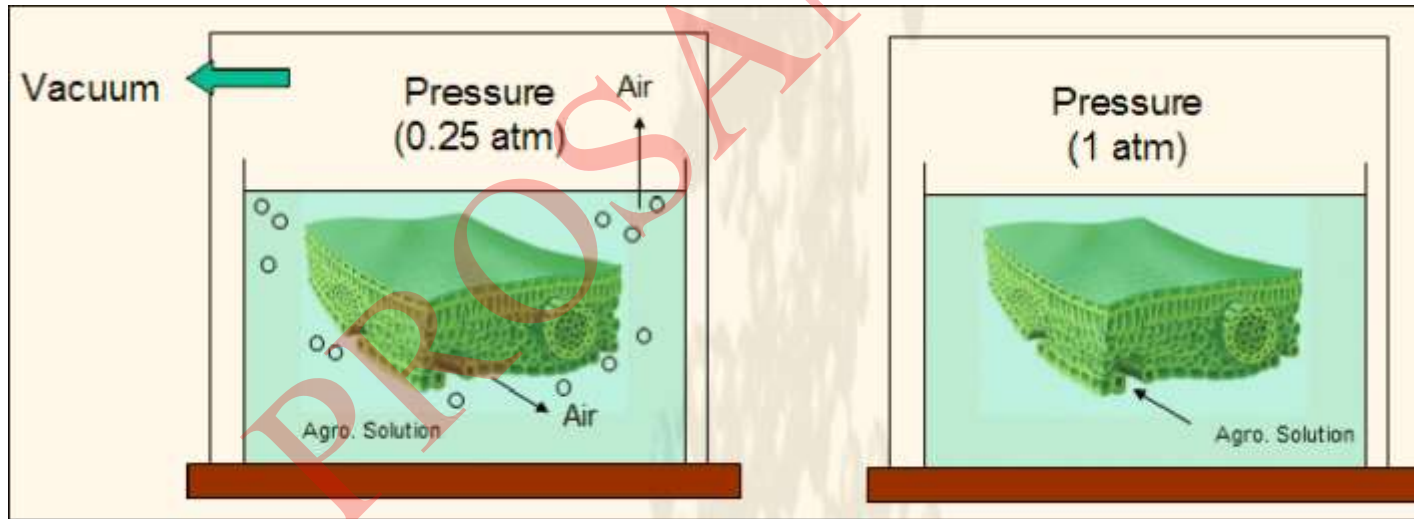
Non-tissue culture based



- ♣ Meristem transformation
- ♣ Floral dip method
- ♣ Pollen transformation

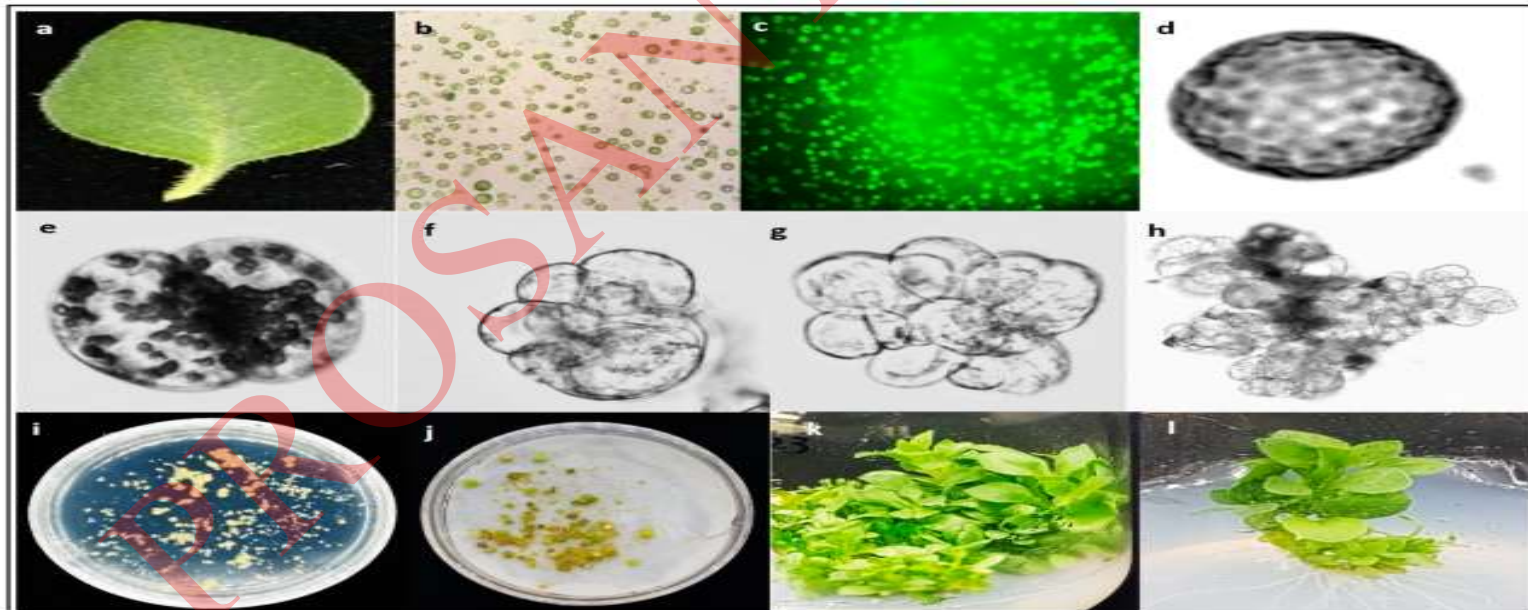
Floral Dip

- Simple submersion of plant into bacterium suspension
- No vacuum is needed
- Conducted with plants grown until just flowering
- Progeny seeds are harvested and germinated using selective antibiotic



Vacuum Infiltration

Protoplast Fusion



Screening Technique

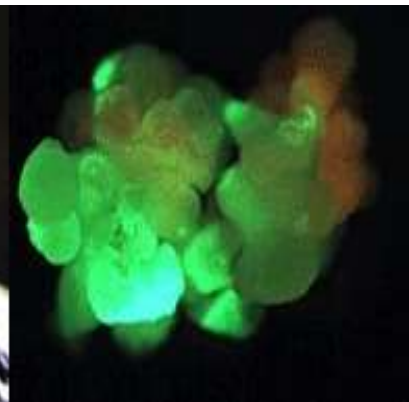
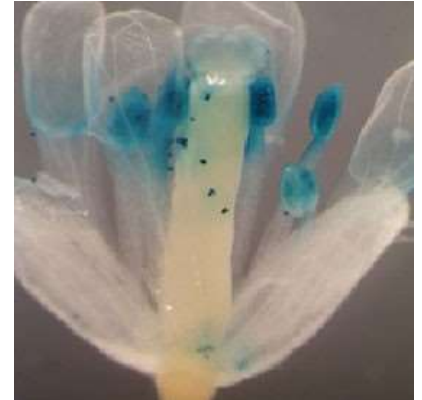
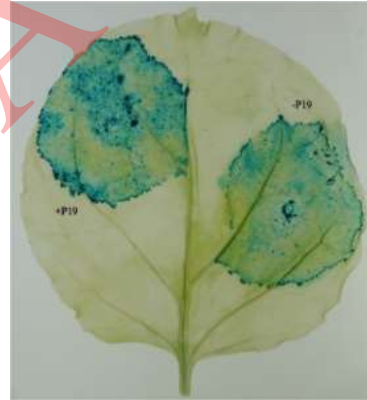
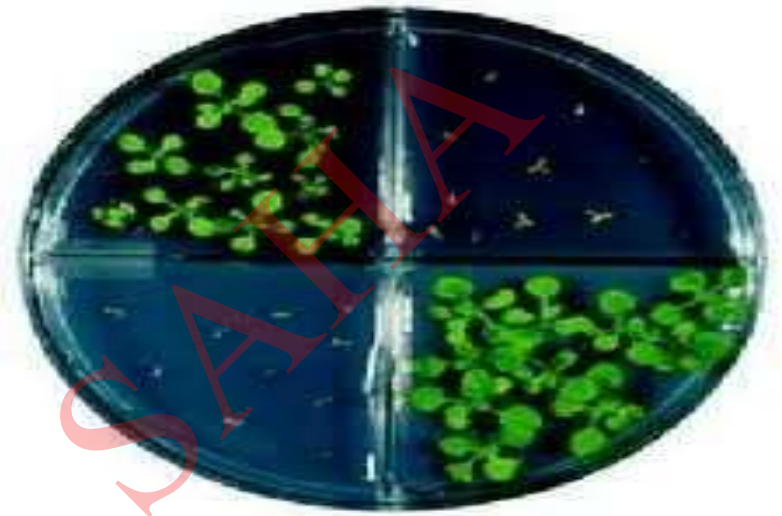
There are many thousands of cells in a leaf disc or callus clump - **only a proportion of these will have taken up the DNA**

therefore can get hundreds of plants back - maybe **only 1% will be transformed**

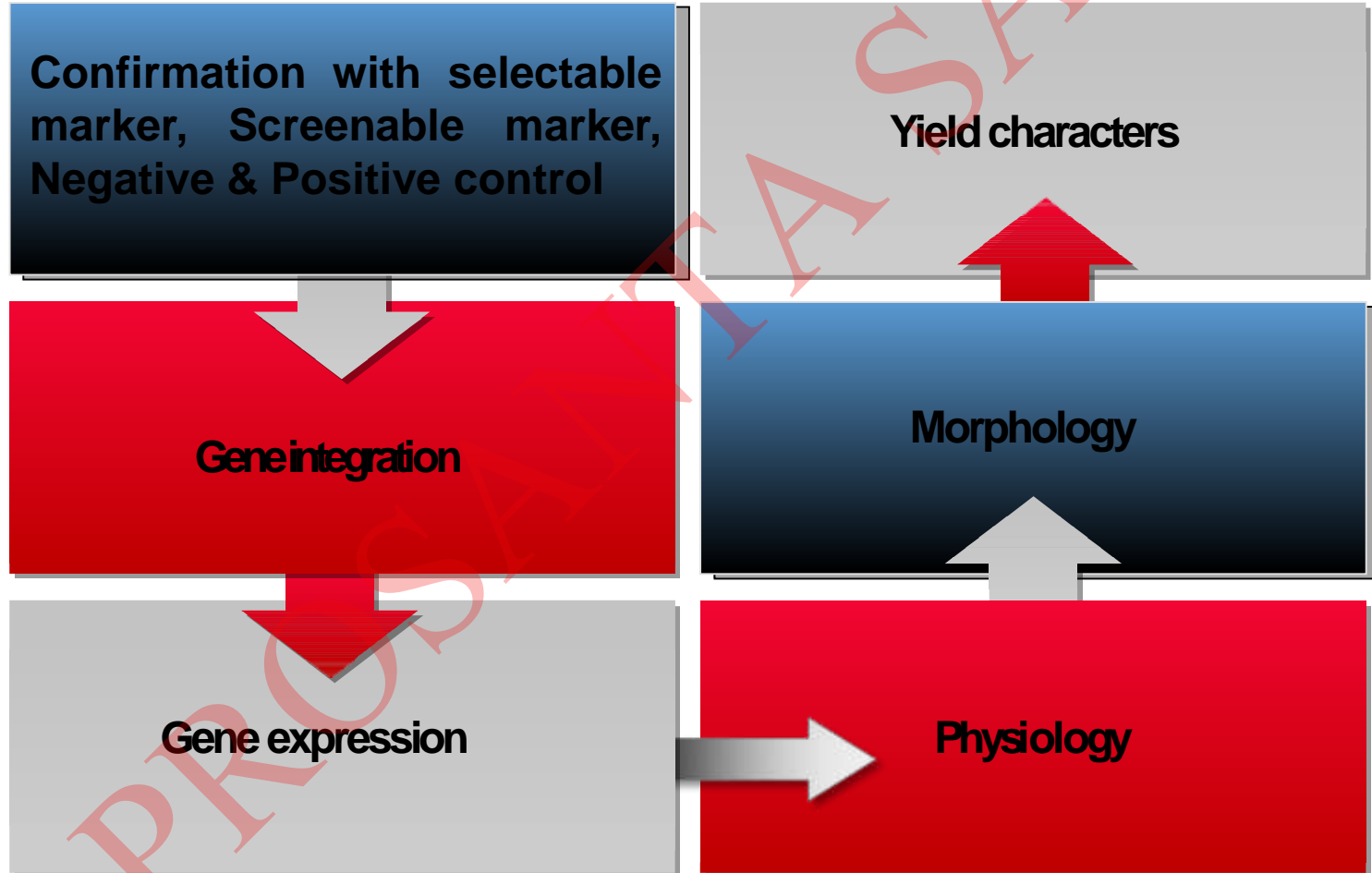
How do we know which plants have taken up the DNA?

Could test each plant - slow, costly

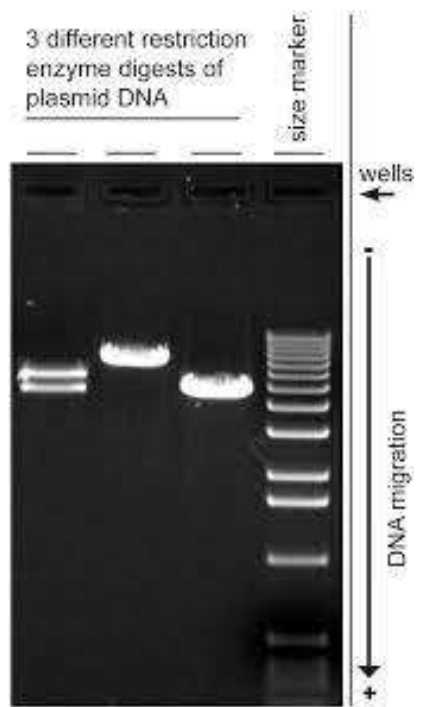
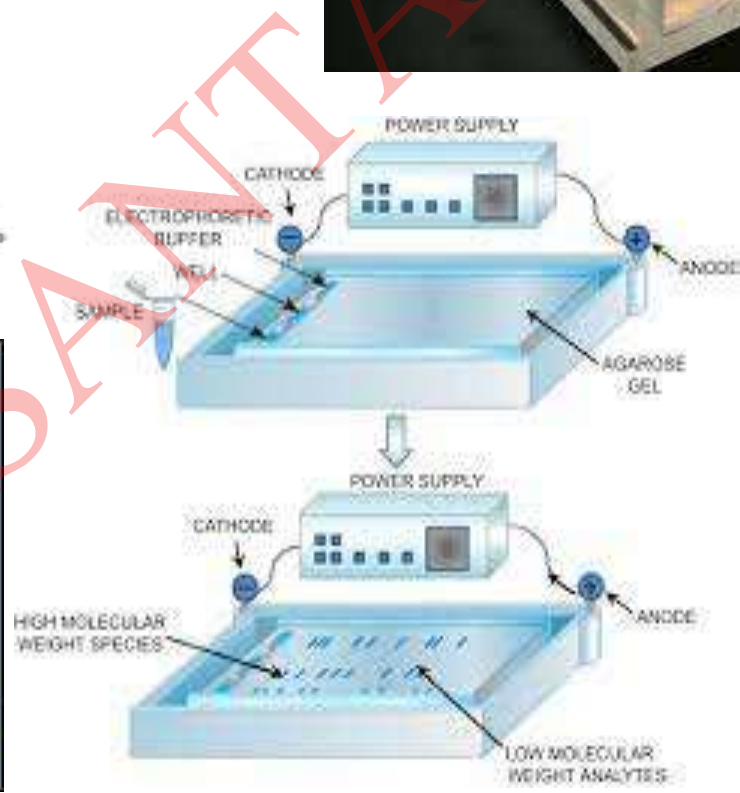
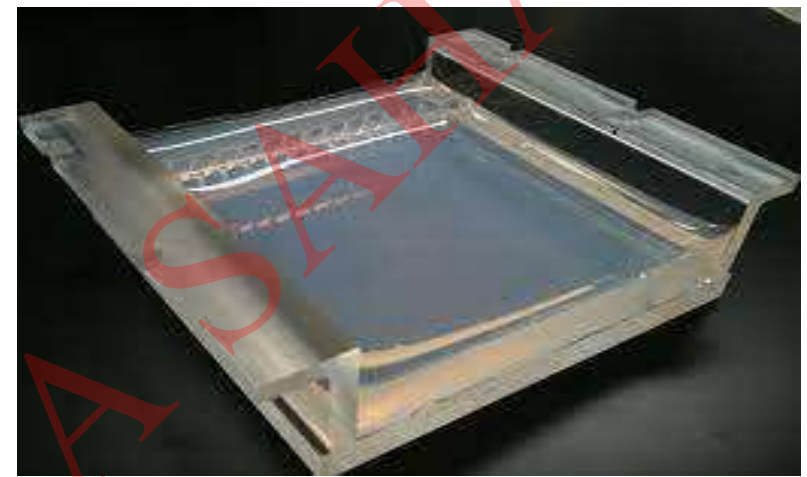
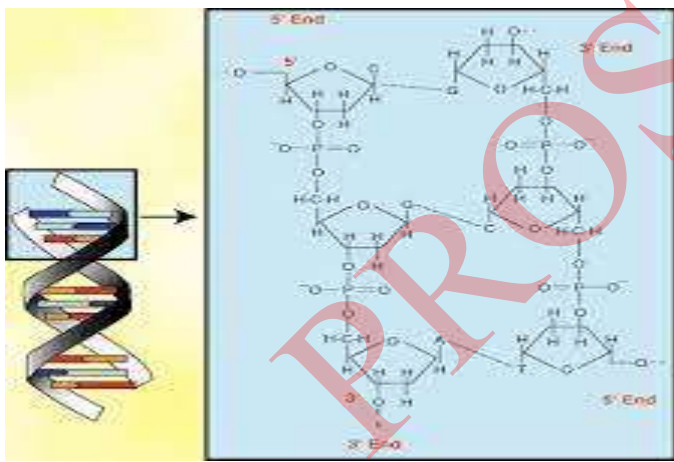
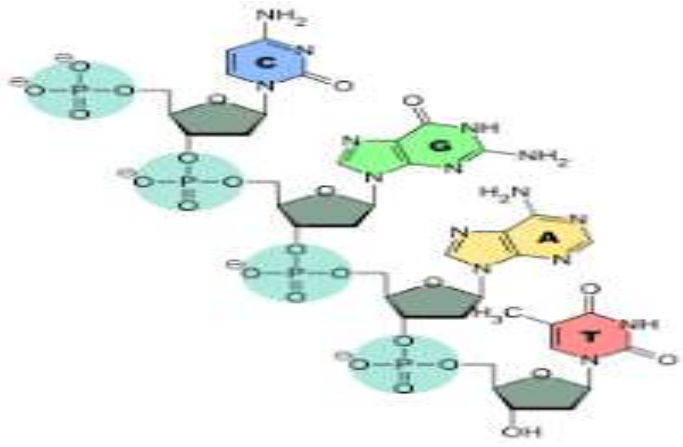
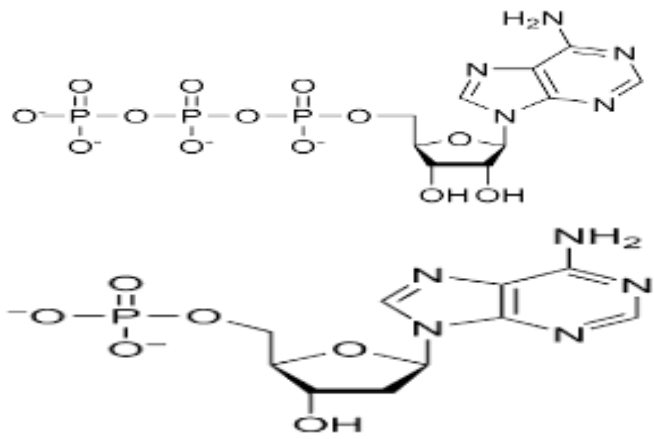
Or use **reporter genes & selectable marker genes**



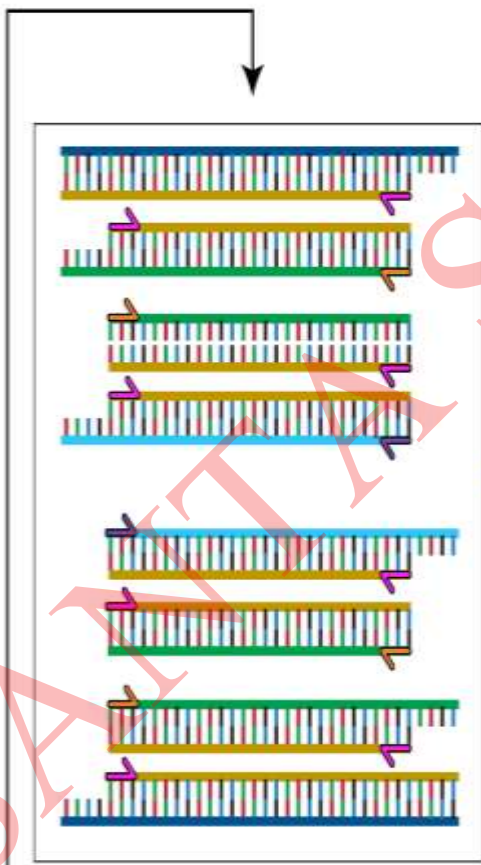
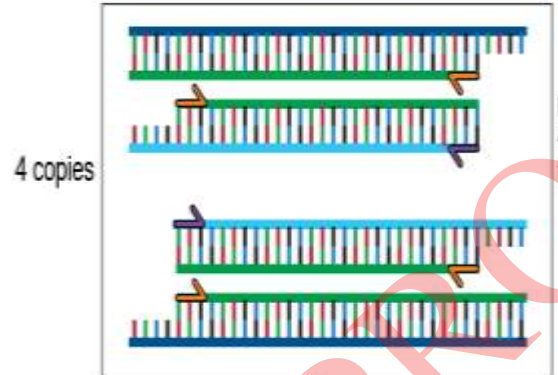
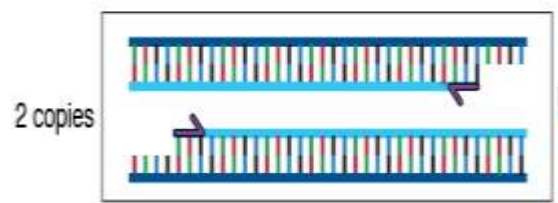
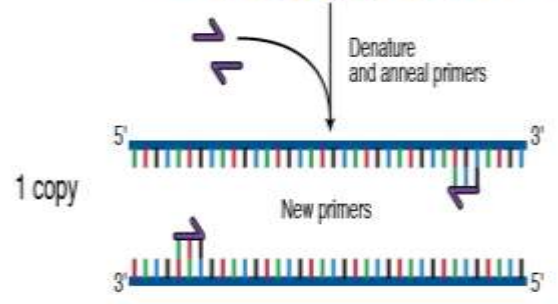
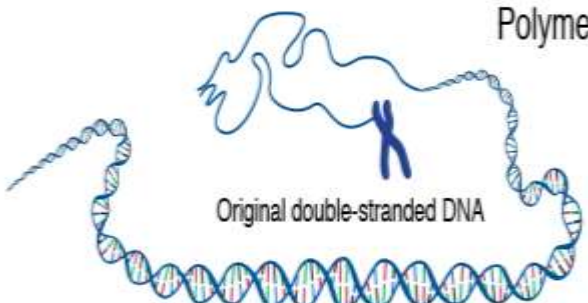
Analysis of T₀ plants



Agarose Gel Electrophoresis)

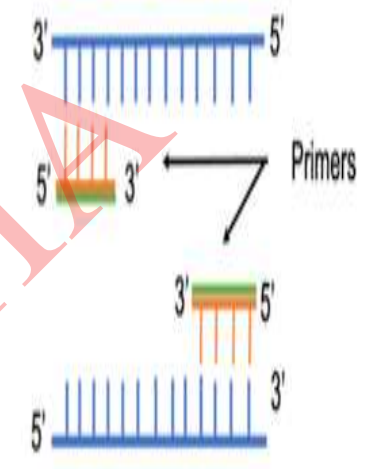


Polymerase Chain Reaction

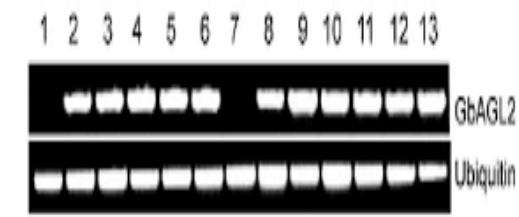


20 -30 cycles

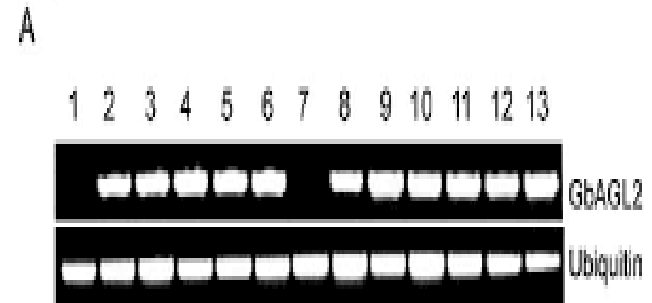
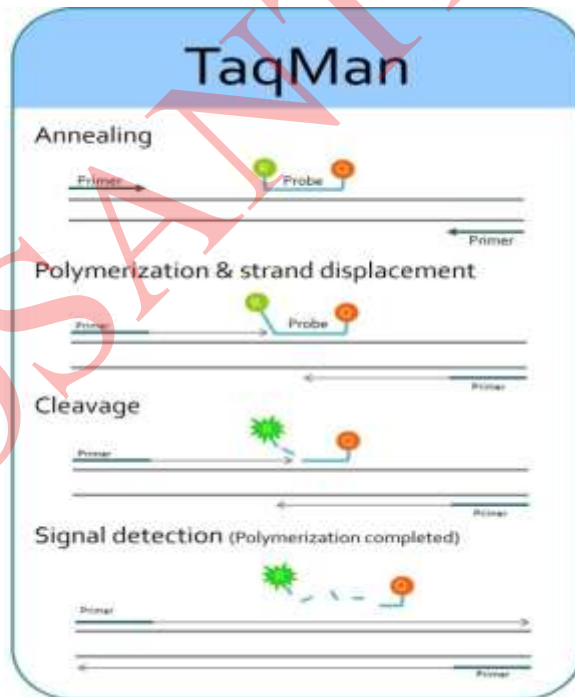
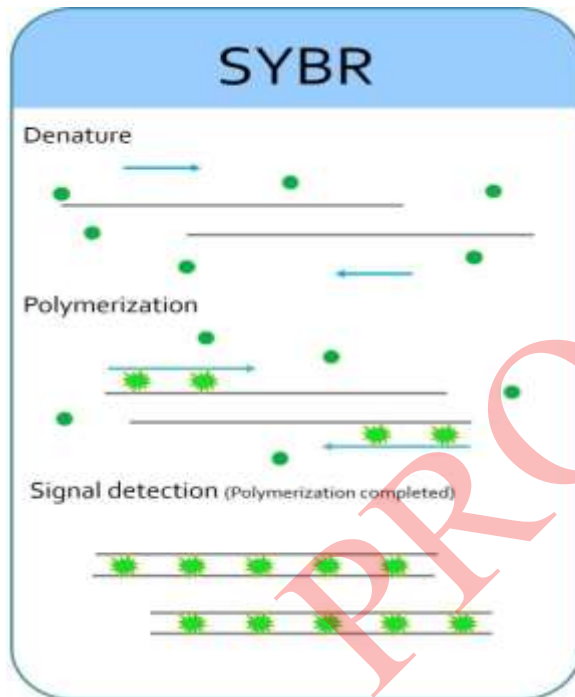
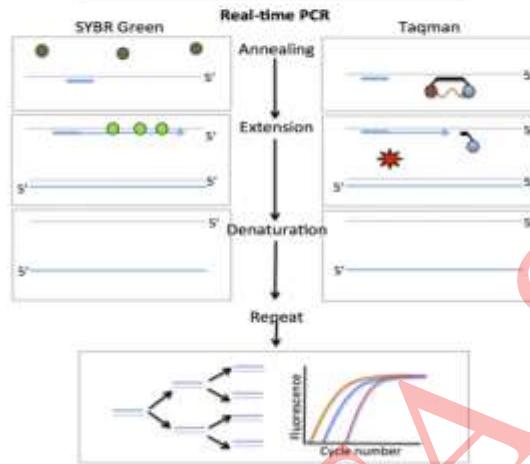
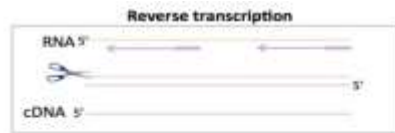
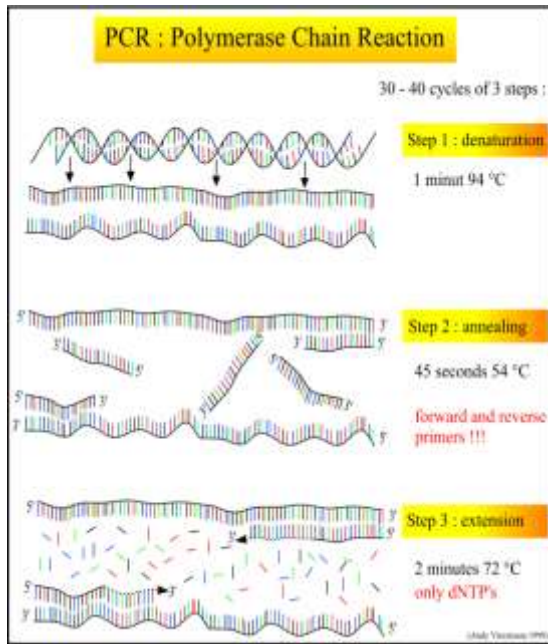
Millions and Millions of copies



A

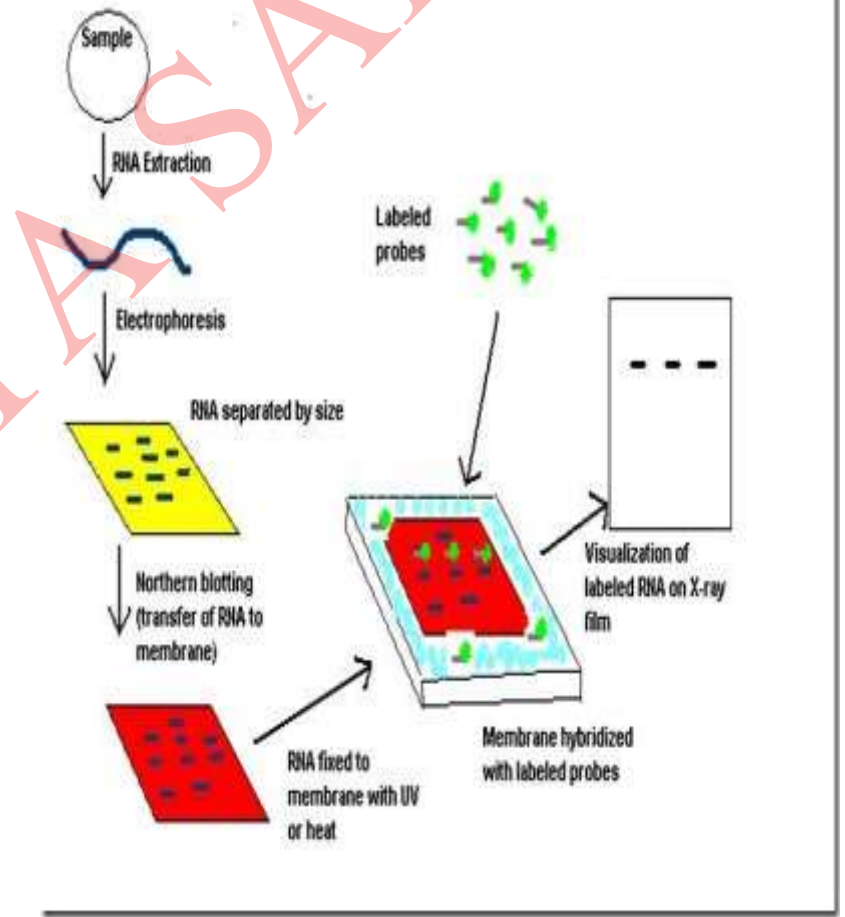
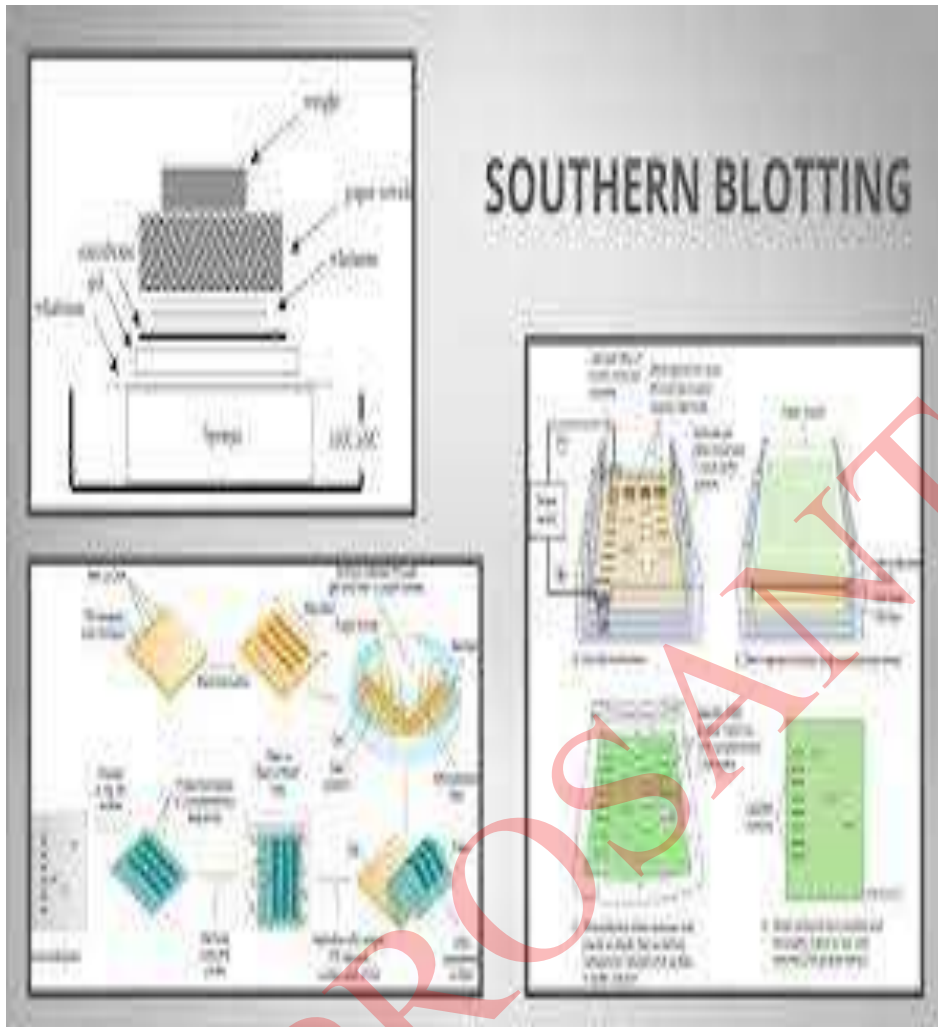


B

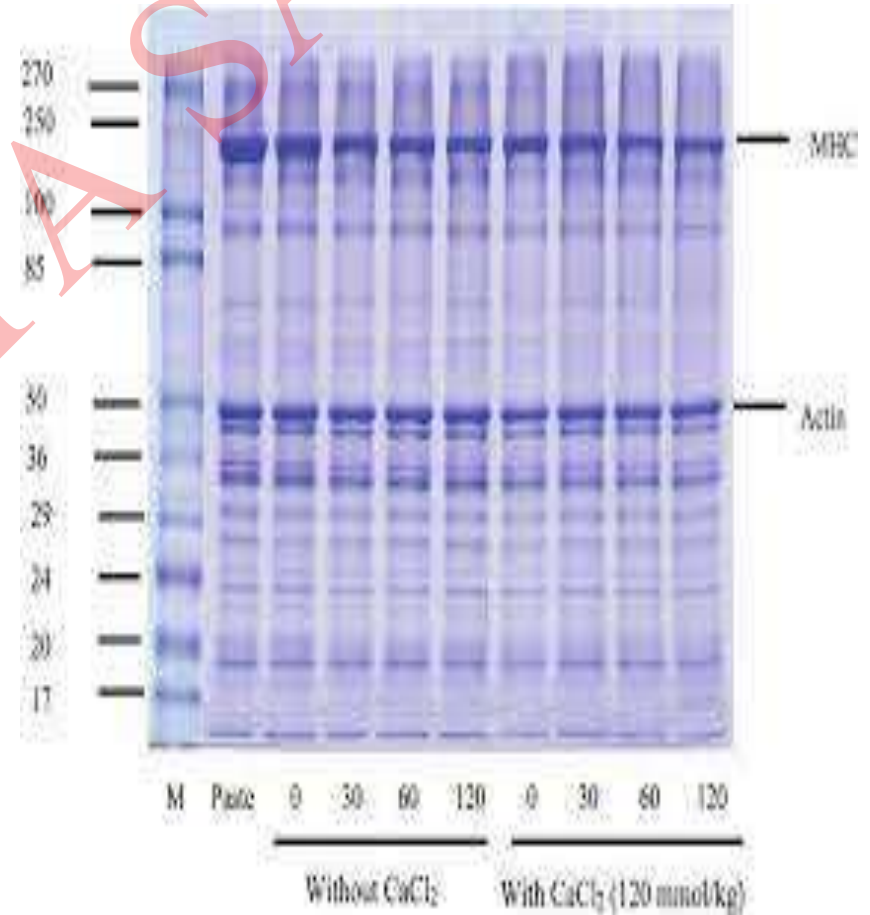
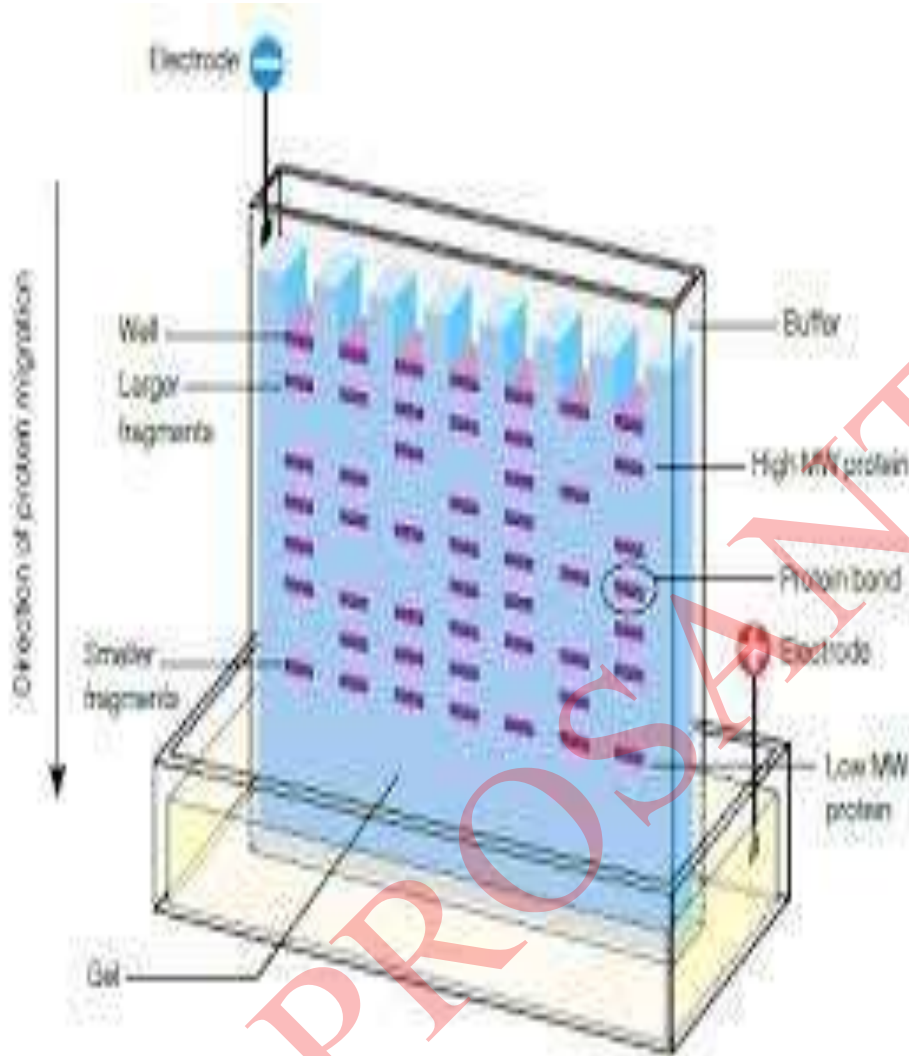


B

Southern and Northern Blotting techniques

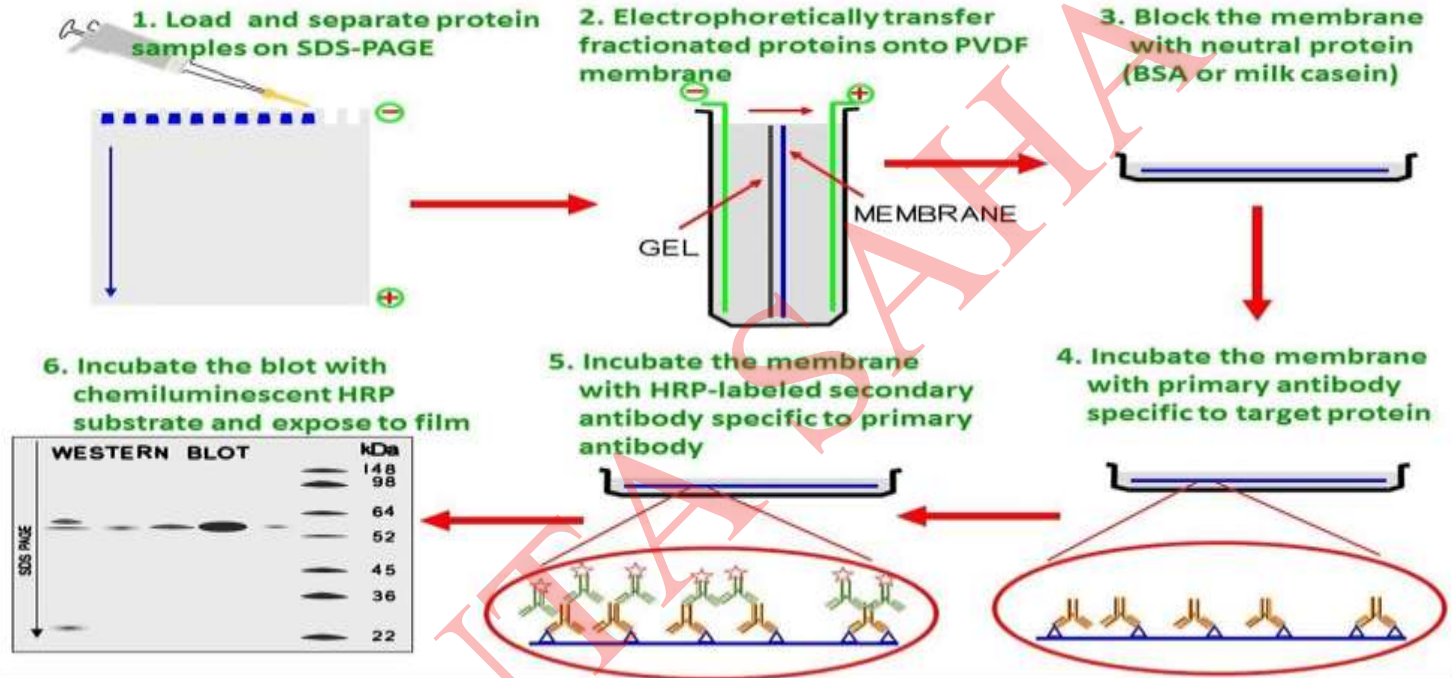


PAGE (Polyacrylamide Gel electrophoresis)



Western Blotting

Western Blotting Procedure



	Southern Blot	Northern Blot	Western Blot
Target molecule	DNA	RNA	Protein
Sample preparation	DNA extraction enzymatic digestion	RNA isolation	Protein extraction
Separation	Electrophoresis	Electrophoresis	Electrophoresis
Membrane material	Nylon	Nylon	Nitrocellulose or PVDF
Probe	Nucleic acid probe with sequence homologous to target	RNA, DNA, or oligodeoxynucleotide	Primary antibody
Probe label	Radiolabel, enzyme	Radiolabel, enzyme	Enzyme
Detection methods	X-ray film, chemiluminescence	X-ray film, chemiluminescence	Film, cooled CCD, camera, LED, or infrared imaging system

Table 1: Comparing Southern, Northern, and Western Blots.

APPLICATIONS

transfer of exogenous genes

Pathogen resistance

Herbicide resistance

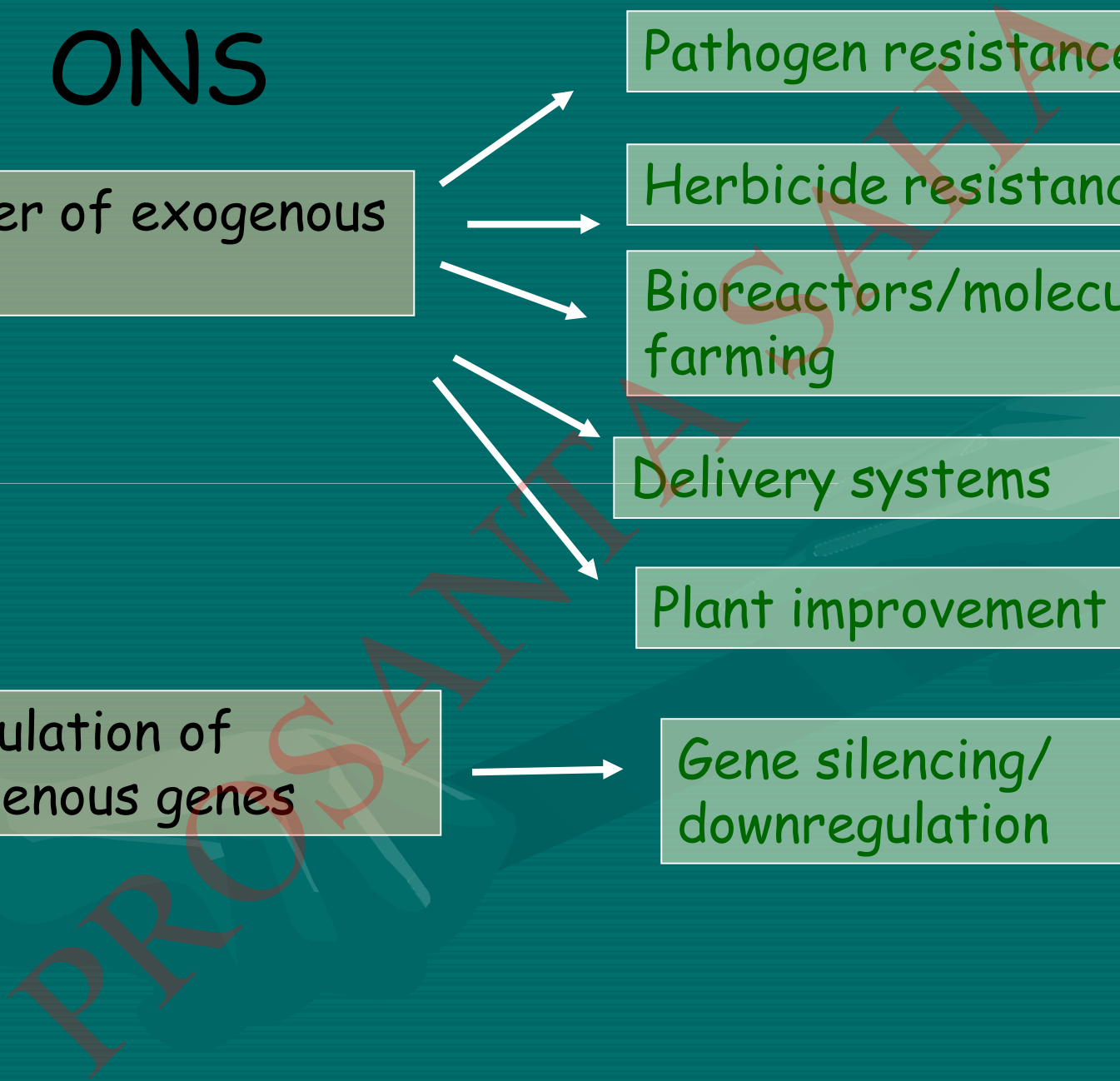
Bioreactors/molecular farming

Delivery systems

Plant improvement

manipulation of endogenous genes

Gene silencing/
downregulation





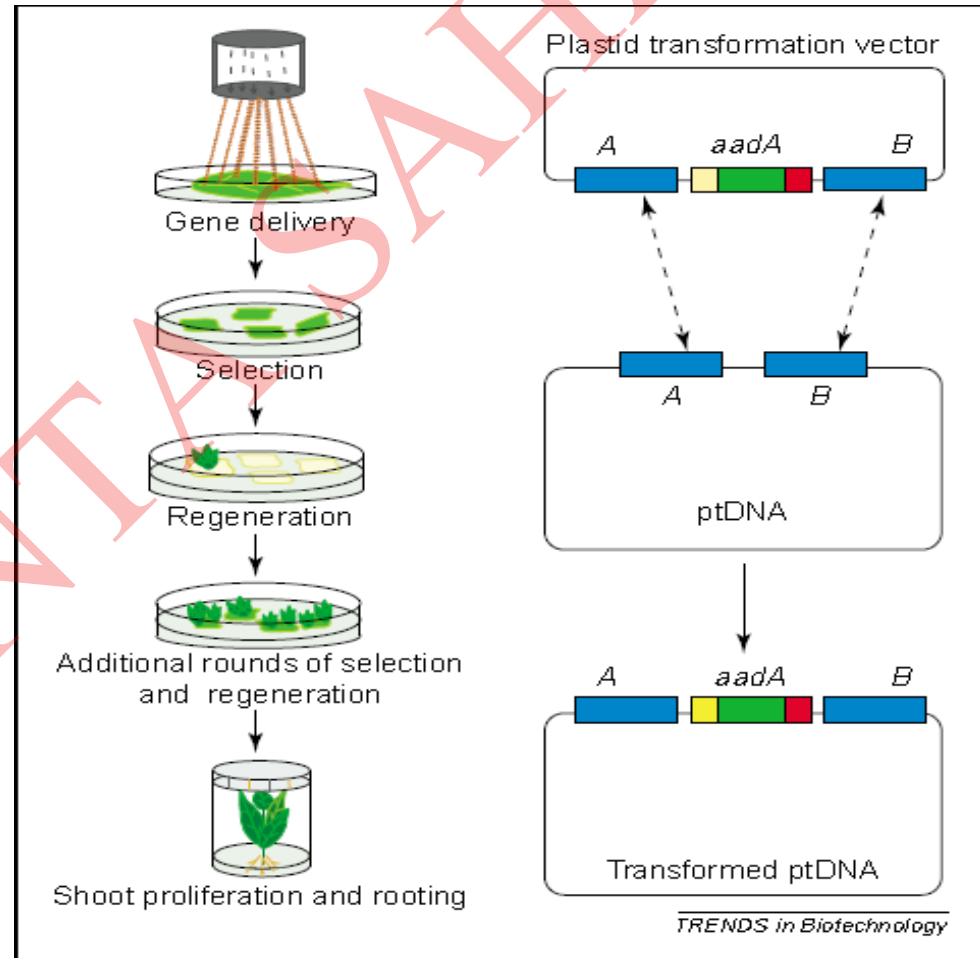
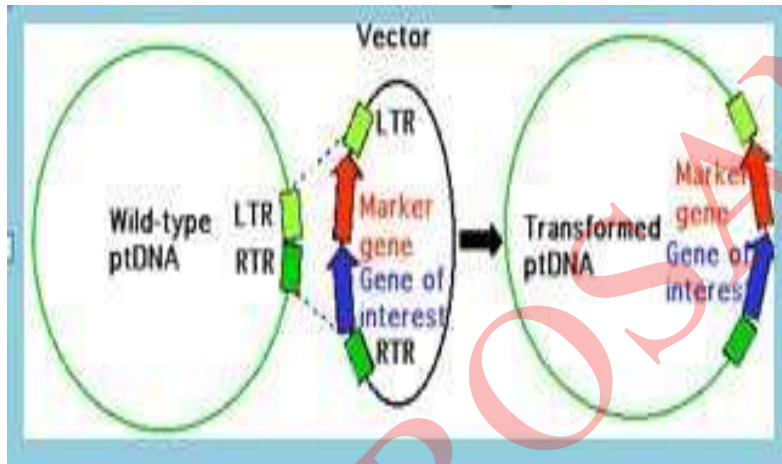
Examples of Genetically modified plants

Table 17.2 Plants that have been genetically transformed

Alfalfa	Carnation	Kiwi fruit	Papaya	Potato	Sunflower
Apple	Carrot	Lettuce	Pea	Red fescue	Sweet potato
<i>Arabidopsis</i>	Corn	Licorice	Peanut	Rice	Tall fescue
Asparagus	Cotton	Lily	Pear	Rye	Tobacco
Banana	Cranberry	Lotus	Pearl millet	Sorghum	Tomato
Barley	Cucumber	Norway spruce	Peony	Soybean	Wheat
Bean	Eggplant	Oat	Petunia	Strawberry	White spruce
Cabbage	Flax	Orchard grass	Plantain	Sugar beet	
Canola	Grape	Orchid	Poplar	Sugarcane	

Chloroplast transformation requires:

1. A chloroplast specific expression vector.
2. A method for DNA delivery through a double membrane of the chloroplast.
3. An efficient selection for the transplastome.



WHAT IS GENE EDITING?



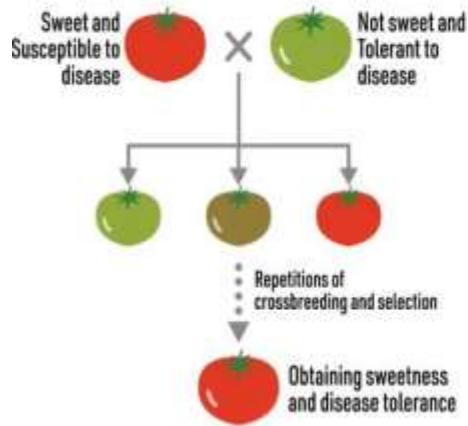
Gene editing is the process of making a tiny, controlled change in the DNA of a living being to produce a GMO.

- Gene editing is used around the world.
- In the United States, federal agencies regulate gene editing.
- Ethicists and concerned parties are carefully debating potential uses for gene editing.

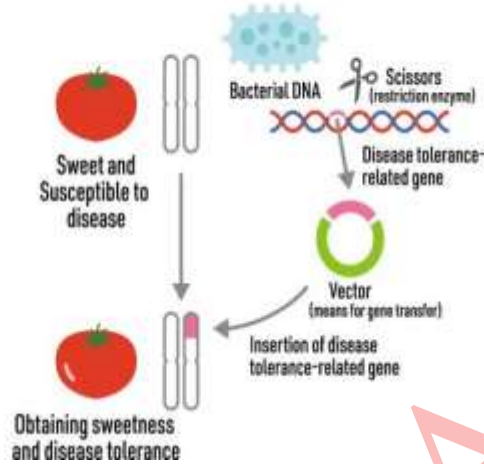


Purdue University is an equal opportunity institution.

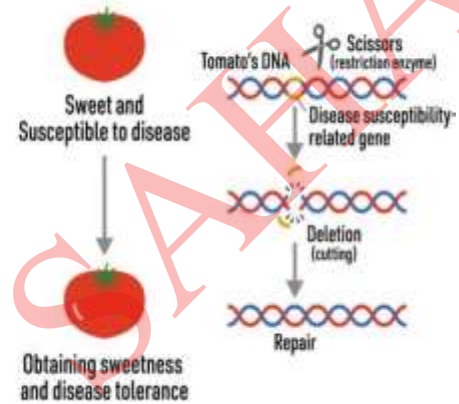
Conventional Breeding



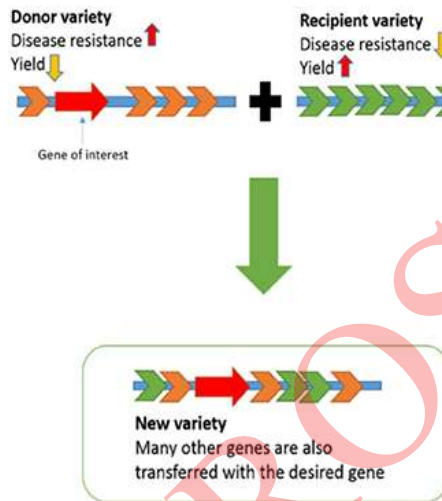
Genetic Modification



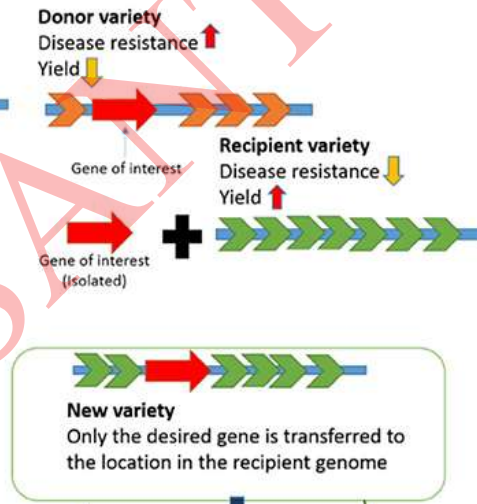
Gene Editing



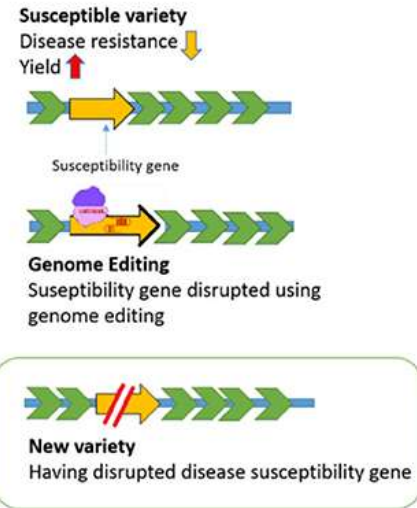
A Conventional Breeding



B Genetic Engineering



C Genome Editing

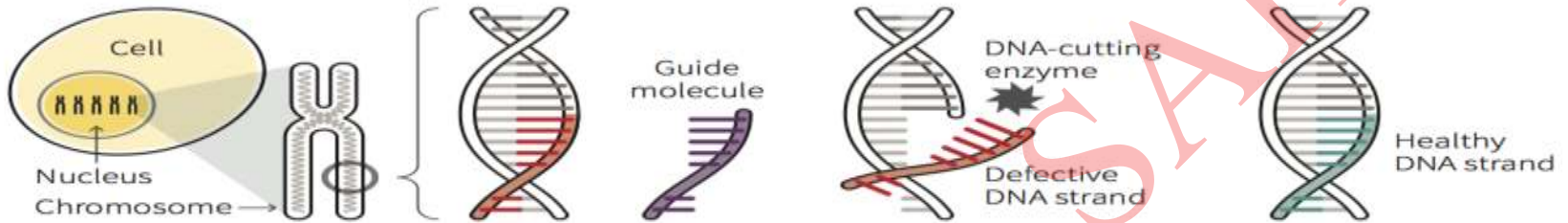


Disease resistance ↑
Yield ↑

DNA editing

A DNA editing technique, called CRISPR/Cas9, works like a biological version of a word-processing programme's "find and replace" function.

HOW THE TECHNIQUE WORKS



A cell is transfected with an enzyme complex containing:

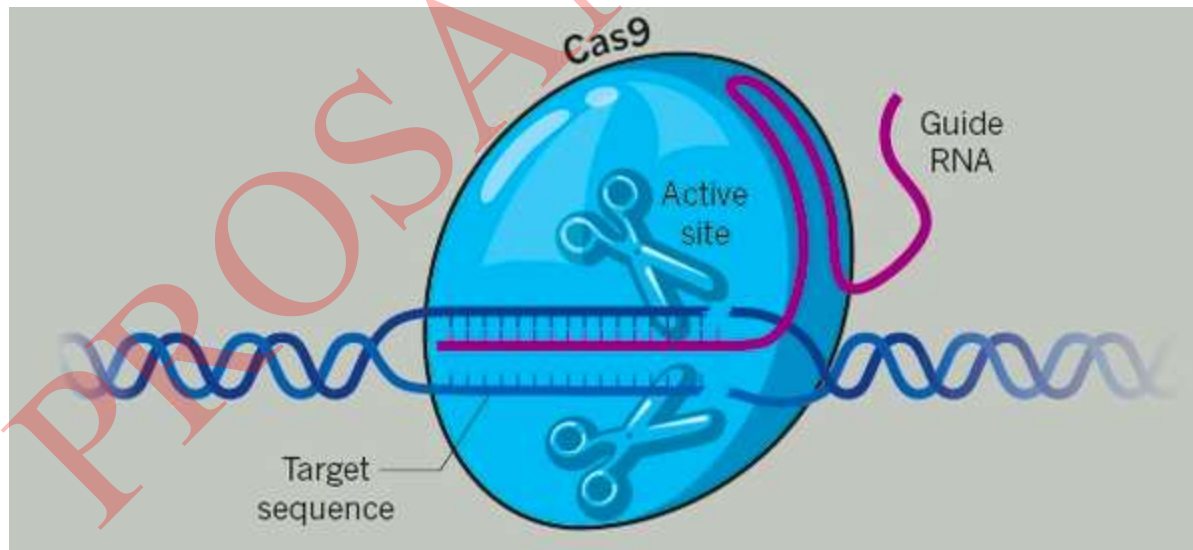
- Guide molecule
- Healthy DNA copy
- DNA-cutting enzyme

A specially designed synthetic guide molecule finds the target DNA strand.

An enzyme cuts off the target DNA strand.

The defective DNA strand is replaced with a healthy copy.

Sources: Reuters; Nature; Massachusetts Institute of Technology



CONCLUSION

Plant breeding techniques result in stable introgression of genes into the genome, but many undesired genes also get transferred..

Alternative is application of plant biotechnology for crop improvement...

Recombinant DNA Technology (RDT) to prepare a construct containing gene of interest and marker sequences under suitable promoters....

Plant tissue culture for *in vitro* regeneration is essential for genetic transformation

Genetic transformation followed by selection result in transfer of only desired genes, but integration into genome is at random location..

Direct method and Vector mediated methods of genetic transformation are operational...

Tissue culture independent methods are also available...

Genome editing techniques modify the genome at only desired locations (Targeted mutation)... And thereby modify the recipient genome to minimum level.....

Thank you!

PROSANTA SAHA